

A supplementary document of

**A general computational model of mitochondrial metabolism  
in a whole organelle scale**

Katsuyuki Yugi and Masaru Tomita

Institute for Advanced Biosciences, Keio University, Japan

# Contents

<b>1</b>	<b>Abbreviations</b>	<b>3</b>
1.1	Species and organs	3
1.2	Metabolites and enzymes: A-G	4
1.3	Metabolites and enzymes: H-Z	5
1.4	Other metabolites	6
<b>2</b>	<b>Reactions</b>	<b>6</b>
2.1	Respiratory chain	6
2.2	TCA cycle	7
2.3	Fatty acid $\beta$ oxidation	7
2.4	Metabolite transporting system	8
<b>3</b>	<b>Parameter classification</b>	<b>8</b>
<b>4</b>	<b>Kinetic parameters</b>	<b>9</b>
4.1	AAC	9
4.2	ACD	10
4.3	ACO	10
4.4	AGC	11
4.5	AlaTA	11
4.6	AspTA	12
4.7	CAC	12
4.8	CIC	13
4.9	Complex I	14
4.10	Complex III	15
4.11	Complex IV	15
4.12	Complex V	16
4.13	CPT I	17
4.14	CPT II	17
4.15	CS	18
4.16	DIC	19
4.17	ECH	19
4.18	ETF-QO	20
4.19	FM	20
4.20	HCD	21
4.21	IDHa	21
4.22	IDHb	22
4.23	MDH	22
4.24	NDK	23
4.25	OCT	24
4.26	OGC	25
4.27	OGDC	26
4.28	PC	26
4.29	PDC	27
4.30	PiC	28
4.31	PYC	29
4.32	SCS	30
4.33	SDH	30

<b>5</b>	<b>Initial condition</b>	<b>31</b>
5.1	The respiratory chain . . . . .	31
5.2	The TCA cycle . . . . .	31
5.3	Fatty acid $\beta$ oxidation . . . . .	32
5.4	The inner-membrane transport . . . . .	33
<b>6</b>	<b>Steady-state condition</b>	<b>34</b>
6.1	The respiratory chain . . . . .	34
6.2	The TCA cycle and the inner-membrane transport . . . . .	35
6.3	Fatty acid $\beta$ oxidation . . . . .	35
<b>7</b>	<b>Rate equations</b>	<b>36</b>
7.1	AAC . . . . .	36
7.2	CB Ordered Bi Bi . . . . .	36
7.3	Complex III . . . . .	37
7.4	Complex V . . . . .	37
7.5	IDHa . . . . .	37
7.6	IDHb . . . . .	37
7.7	Michaelis Uni Uni . . . . .	37
7.8	Multisite Ping-Pong . . . . .	37
7.9	Ordered Bi Bi . . . . .	38
7.10	PC . . . . .	38
7.11	Ping-Pong Bi Bi . . . . .	38
7.12	Rapid Equilibrium Random Bi Bi . . . . .	39
7.13	SCS . . . . .	39
7.14	Uni Uni Reversible . . . . .	39
<b>8</b>	<b>MeSH term and literature search</b>	<b>40</b>

# 1 Abbreviations

## 1.1 Species and organs

Table 1: Abbreviations for species and organs

Abbreviation	Species, Organ
BH	Bovine Heart
BHM	Bovine Heart Mitochondria
BL	Bovine Liver
BLM	Bovine Liver Mitochondria
CL	Chicken Liver
HLC	Human Liver Cytosol
IMS	Intermembrane Space (Mitochondrial)
MAT	Matrix (Mitochondrial)
PH	Pig Heart
PHM	Pig Heart Mitochondria
PL	Pig Liver
PLM	Pig Liver Mitochondria
RbHM	Rabbit Heart Mitochondria
RB	Rat Brain
RH	Rat Heart
RHM	Rat Heart Mitochondria
RK	Rat Kidney
RLM	Rat Liver Mitochondria

## 1.2 Metabolites and enzymes: A-G

Table 2: Abbreviations for metabolites

Abbreviation	Substance name	Compound/EC number
AAC	ATP/ADP Carrier	
ACD	Acyl-CoA Dehydrogenase	EC1.3.99.3
Acetoacetyl-CoA		C00332
Acetyl-CoA		C00024
ACO	Aconitase	EC4.2.1.3
ADP	Adenosine Diphosphate	C00008
AGC	Aspartate/Glutamate Carrier	
Ala	Alanine	C00041
AlaTA	Alanine Transaminase	EC2.6.1.2
Asp	Aspartate	C00049
AspTA	Aspartate Transaminase	EC2.6.1.1
ATP	Adenosine Triphosphate	C00002
CAC	Carnitine Carrier	
Car	Carnitine	C00318
CIC	Citrate Carrier	
Cit	Citrate	C00158
CPT-I	Carnitine Palmitoyl Transferase I	EC2.3.1.21
CPT-II	Carnitine Palmitoyl Transferase II	EC2.3.1.21
CoA	Coenzyme A	C00010
Complex-I	NADH Dehydrogenase	EC1.6.5.3
Complex-III	Ubiquinol:Cytochrome c Oxidoreductase	EC1.10.2.2
Complex-IV	Cytochrome c Oxidase	EC1.9.3.1
Complex-V	ATP Synthetase	EC3.6.1.34
CO <sub>2</sub>	Carbon Dioxide	C00011
CS	Citrate Synthase	EC4.1.3.7
Cyt-c <sub>2+</sub>	Ferricytochrome c	C00125
Cyt-c <sub>3+</sub>	Ferrocycytochrome c	C00126
DIC	Dicarboxyrate Carrier	
ECH	Enoyl-CoA Hydratase	EC4.2.1.17
ETF <sub>ox</sub>	Electron Transfer Flavoprotein (oxidised form)	
ETF <sub>red</sub>	Electron Transfer Flavoprotein (reduced form)	
ETF-QO	ETF:Q Oxidoreductase	
FM	Fumarase	EC4.2.1.2
Fum	Fumarate	C00122
GDP	Guanosine Diphosphate	C00035
Glu	Glutamate	C00025
GTP	Guanosine Triphosphate	C00044

### 1.3 Metabolites and enzymes: H-Z

Table 3: Abbreviations for metabolites (cont'd)

Abbreviation	Substance name	Compound/EC number
HCD	Hydroxyacyl-CoA Dehydrogenase	EC1.1.1.35
H+	Hydrogen ion (proton)	C00080
IDHa	Isocitrate Dehydrogenase (NAD <sup>+</sup> )	EC1.1.1.41
IDHb	Isocitrate Dehydrogenase (NADP <sup>+</sup> )	EC1.1.1.42
IsoCit	Isocitrate	C00311
Mal	Malate	C00149
MDH	Malate Dehydrogenase	EC1.1.1.37
NAD <sup>+</sup>		C00003
NADH		C00004
NADP <sup>+</sup>		C00006
NADPH		C00005
NDK	Nucleoside Diphosphate Kinase	EC2.7.4.6
OCT	Oxoacyl-CoA Thiolase	EC2.3.1.16
OG	Oxoglutarate	C00026
OGC	Oxoglutarate Carrier	
OGDC	Oxoglutarate Dehydrogenase Complex	EC1.2.4.2 etc.
OXA	Oxaloacetate	C00036
PalCar	Palmitoylcarnitine	C02990
PC	Pyruvate Carboxylase	EC6.4.1.1
PDC	Pyruvate Dehydrogenase Complex	EC1.2.4.1 etc.
Pi	Phosphate	C00009
PiC	Pi Carrier	
Pyr	Pyruvate	C00022
PYC	Pyruvate Carrier	
Q	Ubiquinone	C00399
QH2	Ubiquinol	C00390
SCoA	Succinyl-CoA	C00091
SCS	Succinyl-CoA synthetase	EC6.2.1.4
SDH	Succinate Dehydrogenase	EC1.3.5.1
Suc	Succinate	C00042

## 1.4 Other metabolites

Table 4: Abbreviations for metabolites (cont'd)

Abbreviation	Substance name	Compound/EC number
10Acyl-CoA	Decanoyl-CoA	C05274
10Enoyl-CoA	trans-Dec-2-enoyl-CoA	C05275
10Hydroxyacyl-CoA	(S)-3-Hydroxydedecanoyl-CoA	C05264
10Oxoacyl-CoA	3-Oxodecanoyl-CoA	C05265
12Acyl-CoA	Lauroyl-CoA	C01832
12Enoyl-CoA	trans-Dodec-2-enoyl-CoA	C03221
12Hydroxyacyl-CoA	(S)-3-Hydroxydodecanoyl-CoA	C05262
12Oxoacyl-CoA	3-Oxododecanoyl-CoA	C05263
14Acyl-CoA	Myristoyl-CoA	C02593
14Enoyl-CoA	trans-Tetradec-2-enoyl-CoA	C05273
14Hydroxyacyl-CoA	(S)-3-Hydroxytetradecanoyl-CoA	C05260
14Oxoacyl-CoA	3-Oxotetradecanoyl-CoA	C05261
16Acyl-CoA	Palmitoyl-CoA	C00154
16Enoyl-CoA	trans-Hexadec-2-enoyl-CoA	C05272
16Hydroxyacyl-CoA	(S)-3-Hydroxyhexadecanoyl-CoA	C05258
16Oxoacyl-CoA	3-Oxohexadecanoyl-CoA	C05259
4Acyl-CoA	Butanoyl-CoA	C00136
4Enoyl-CoA	Crotonyl-CoA	C00877
4Hydroxyacyl-CoA	(S)-3-Hydroxybutanoyl-CoA	C01144
6Acyl-CoA	Hexanoyl-CoA	C05270
6Enoyl-CoA	trans-Hex-2-enoyl-CoA	C05271
6Hydroxyacyl-CoA	(S)-3-Hydroxyhexanoyl-CoA	C05268
6Oxoacyl-CoA	3-Oxohexanoyl-CoA	C05269
8Acyl-CoA	Octanoyl-CoA	C01944
8Enoyl-CoA	trans-Oct-2-enoyl-CoA	C05276
8Hydroxyacyl-CoA	(S)-3-Hydroxyoctanoyl-CoA	C05266
8Oxoacyl-CoA	3-Oxoocatanoyl-CoA	C05267

## 2 Reactions

### 2.1 Respiratory chain

Table 5: Reactions in the respiratory chain (where  $H_{MAT}^+$  denotes  $H^+$  in the matrix,  $H_{IMS}^+$  is  $H^+$  in the intermembrane space)

complex	reaction	reaction mechanism	source
I	$NADH + Q + 5H_{MAT}^+ \longleftrightarrow NAD^+ + QH_2 + 4H_{IMS}^+$	Ping-Pong Bi Bi [Fato et al., 1996]	BHM
II(SDH)	$Suc + Q \longleftrightarrow Fum + QH_2$	Ping-Pong Bi Bi [Grivennikova et al., 1993]	BHM
III	$QH_2 + 2cyt\ c^{3+} + 2H_{MAT}^+ \rightarrow Q + 2cyt\ c^{2+} + 4H_{IMS}^+$	See [Kubota et al., 1992]	BHM
IV	$4cyt\ c^{2+} + O_2 + 8H_{MAT}^+ \rightarrow 4cyt\ c^{3+} + 2H_2O + 4H_{IMS}^+$	Michaelis Uni Uni [Malmström and Andréasson, 1985]	-
V	$ADP + Pi + 3H_{IMS}^+ \longleftrightarrow ATP + H_2O + 3H_{MAT}^+$	See [Kholodenko, 1993]	-

## 2.2 TCA cycle

Table 6: The enzymes in/around the TCA cycle

enzyme	reaction	reaction mechanism	source
PDC	$\text{Pyr} + \text{NAD}^+ + \text{CoA} \longrightarrow \text{Acetyl-CoA} + \text{NADH} + \text{CO}_2$	See [Hamada et al., 1975]	PHM
PC	$\text{Pyr} + \text{ATP} + \text{CO}_2 \longleftrightarrow \text{OXA} + \text{ADP} + \text{Pi}$	See [Barden et al., 1972]	CL
AspTA	$\text{Asp} + \text{OG} \longleftrightarrow \text{OXA} + \text{Glu}$	Ping-Pong Bi Bi [Velick and Vavra, 1962, Henson and Cleland, 1964]	PH
AlaTA	$\text{Ala} + \text{OG} \longleftrightarrow \text{Glu} + \text{Pyr}$	Ping-Pong Bi Bi [De Rosa et al., 1979]	PL
NDC	$\text{ATP} + \text{GDP} \longleftrightarrow \text{ADP} + \text{GTP}$	Ping-Pong Bi Bi [Garces and Cleland, 1969]	yeast
CS	$\text{OXA} + \text{Acetyl-CoA} \longleftrightarrow \text{Cit} + \text{CoA}$	Random Bi Bi [Shepherd and Garland, 1969, Matsuoka and Srere, 1973]	RK, RB
ACO	$\text{Cit} \longleftrightarrow \text{IsoCit}$	Uni Uni Reversible [Guarriero-Bobyleva et al., 1978]	RLM
IDHa	$\text{IsoCit} + \text{NAD}^+ \longrightarrow \text{OG} + \text{NADH}$	See [Plaut et al., 1974]	BH
IDHb	$\text{IsoCit} + \text{NADP}^+ \longleftrightarrow \text{OG} + \text{NADPH}$	See [Londesborough and Dalziel, 1970]	BHM
OGDC	$\text{OG} + \text{NAD}^+ + \text{CoA} \longrightarrow \text{SCoA} + \text{NADH} + \text{CO}_2$	See [Hamada et al., 1975]	PHM
SCS	$\text{SCoA} + \text{GDP} + \text{Pi} \longleftrightarrow \text{Suc} + \text{CoA} + \text{GTP}$	See [Cha and Parks Jr., 1964]	PH
SDH	$\text{Suc} + \text{Q} \longleftrightarrow \text{Fum} + \text{QH}_2$	Ping-Pong Bi Bi [Grivennikova et al., 1993]	BHM
FM	$\text{Fum} \longleftrightarrow \text{Mal}$	Uni Uni Reversible	
MDH	$\text{Mal} + \text{NAD}^+ \longleftrightarrow \text{OXA} + \text{NADH}$	Ordered Bi Bi [Crow et al., 1983]	HLC

## 2.3 Fatty acid $\beta$ oxidation

Table 7: The enzymes in the fatty acid  $\beta$  oxidation

enzyme	reaction	reaction mechanism	source
ACD	$\text{Acyl-CoA} + \text{ETF}_{ox} \longleftrightarrow \text{Enoyl-CoA} + \text{ETF}_{red}$	Ordered Bi Bi [McKean et al., 1979]	PLM
ECH	$\text{Enoyl-CoA} + \text{H}_2\text{O} \longleftrightarrow \text{3-hydroxyacyl-CoA}$	Uni Uni Reversible [Yang and Schulz, 1987]	BL
HCD	$\text{3-hydroxyacyl-CoA} + \text{NAD}^+ \longrightarrow \text{3-oxoacyl-CoA} + \text{NADH}$	Michaelis Uni Uni [Yang and Schulz, 1987]	PH
OCT	$\text{3-oxoacyl-CoA} + \text{CoA} \longleftrightarrow \text{Acyl-CoA} + \text{Acetyl-CoA}$	Ping-Pong Bi Bi [Miyazawa et al., 1981]	RLM
ETF-QO	$\text{ETF}_{red} + \text{Q} \longleftrightarrow \text{ETF}_{ox} + \text{QH}_2$	Ping-Pong Bi Bi [Beckmann and Frerman, 1985]	PLM
CPT I	$\text{16Acyl-CoA} + \text{Car} \longleftrightarrow \text{CoA} + \text{PalCar}$	Rapid Equilibrium Random Bi Bi [Ramsay et al., 1987]	BLM
CPT II	$\text{CoA} + \text{PalCar} \longleftrightarrow \text{16Acyl-CoA} + \text{Car}$	Ordered Bi Bi [Mann et al., 1995]	RLM
CAC	$\text{PalCar}_{IMS} + \text{Car}_{MAT} \longleftrightarrow \text{PalCar}_{MAT} + \text{Car}_{IMS}$	Ping-Pong Bi Bi [Indiveri et al., 1994]	RLM



## 2.4 Metabolite transporting system

Table 8: Metabolite carriers on the inner membrane

enzyme	reaction	reaction mechanism	source
AAC	$ATP_{MAT} \longrightarrow ATP_{IMS}$ $ADP_{MAT} \longleftarrow ADP_{IMS}$	See [Krämer and Klingenberg, 1982]	RHM
PiC	$Pi_{IMS} + H_{IMS}^+$ $\longleftrightarrow Pi_{MAT} + H_{MAT}^+$	Rapid Equilibrium Random Bi Bi [Stappen and Krämer, 1994]	RHM
PYC	$Pyr_{IMS} + H_{MAT}^+$ $\longleftrightarrow Pyr_{MAT} + H_{IMS}^+$	Rapid Equilibrium Random Bi Bi ("Sequential Mechanism" [Nalecz, 1994])	RLM
OGC	$OG_{IMS} + Mal_{MAT}$ $\longleftrightarrow OG_{MAT} + Mal_{IMS}$	Rapid Equilibrium Random Bi Bi [Indiveri et al., 1991a]	BHM
DIC	$Mal_{IMS} + Pi_{MAT}$ $\longleftrightarrow Mal_{MAT} + Pi_{IMS}$	Rapid Equilibrium Random Bi Bi [Indiveri et al., 1993]	RLM
CIC	$Cit_{IMS} + Mal_{MAT}$ $\longleftrightarrow Cit_{MAT} + Mal_{IMS}$	Rapid Equilibrium Random Bi Bi [Bisaccia et al., 1993]	RLM
AGC	$Asp_{IMS} + Glu_{MAT}$ $\longleftrightarrow Asp_{MAT} + Glu_{IMS}$	Rapid Equilibrium Random Bi Bi [Sluse et al., 1991]	RHM
CAC	$PalCar_{IMS} + Car_{MAT}$ $\longleftrightarrow PalCar_{MAT} + Car_{IMS}$	Ping-Pong Bi Bi [Indiveri et al., 1994]	RLM

## 3 Parameter classification

We classified all the kinetic parameters into four classes to distinguish their background as follows. This classification rule was applied for annotating the parameters shown in 4.1 ~ 4.33.

Table 9: The four classes for annotating the kinetic parameters

Class	Definition	Example
class 0	Found in the literature	$K_{mA} = 2.3E - 3(M)$ , $K_{mB} = 2.3 \pm 0.2E - 3(M)$
class 1	Estimated around the values in the literature	$K_m = 2.3E - 3(M)$ $\longrightarrow K_m = 2.6E - 3(M)$
class 2	Estimated around the values of analogous metabolites	$K_{mATP} = 2.3E - 3(M)$ $\longrightarrow 0 \leq K_{mGTP} \leq 3E - 3(M)$
class 3	Estimated arbitrarily	$? \leq k \leq ?$ $\longrightarrow k = 1.2E + 9 \text{ sec}^{-1}$

## 4 Kinetic parameters

### 4.1 AAC

Table 10: Kinetic parameters and their sources(AAC)

Parameter	class	notice
kf0	0.9	class 0
kr0	0.9	class 0
normalize	2.21	class 0
Kd1	5.9E-4	class 3
Kd2	5.9E-4	class 3
Cf	3.30	class 0
Cr	-3.34	class 0
T	310.0	-
kinetic mechanism rate equation source for parameter estimation		see [Krämer and Klingenberg, 1982] See 7.1 [Krämer and Klingenberg, 1982] Figure 2(B) $V_{\rightarrow}^D(\Delta\Psi = 0, 180\text{mV})$

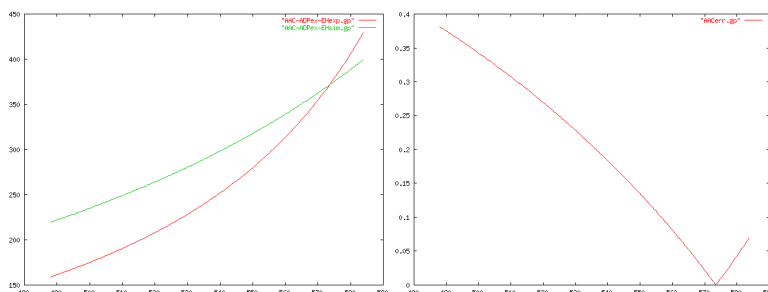


Figure 1: Comparison between experimental data and computed ones from estimated parameters(AAC)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.2 ACD

Table 11: Kinetic parameters and their sources(ACD)

Parameter		class	notice
KmS1	39E-6	class 0	[McKean et al., 1979, Table 1]
KmS2	0.12E-6	class 0	
KmP1	1.08E-6	class 2	
KmP2	2.42E-5	class 2	
KiS1	76E-6	class 0	
KiS2	0.24E-6	class 0	
KiP1	7.53E-5	class 2	
KiP2	1.19E-5	class 2	
Keq	8.99	class 3	
KcF	2.18	class 0	
KcR	0.30	class 2	
kinetic mechanism			Ordered Bi Bi
rate equation			[McKean et al., 1979]
source for parameter estimation			See 7.9
			[McKean et al., 1979]

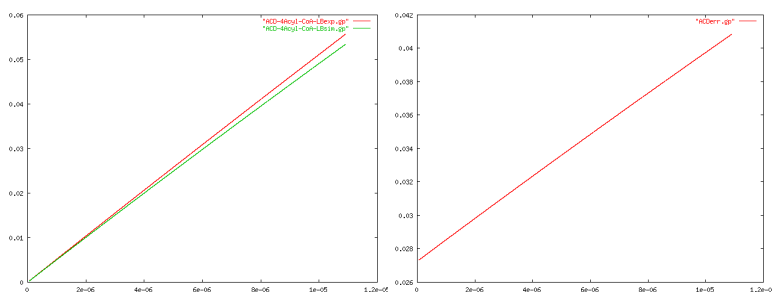


Figure 2: Comparison between experimental data and Computed data from estimated parameters(ACD)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.3 ACO

Table 12: Kinetic parameters and their sources(ACO)

Parameter		class	notice
Ks	0.50E-3	class 0	calculated from the graph
Kp	0.11E-3	class 0	
KcF	20.47	class 0	
KcR	31.44	class 0	
kinetic mechanism			Uni Uni Reversible
rate equation			[Guarriero-Bobyleva et al., 1978]
			See 7.14

## 4.4 AGC

Table 13: Kinetic parameters and their sources(AGC)

Parameter		class	notice
KiS1	80E-6	class 0	[Dierks and Krämer, 1988]
KiS2	3.2E-3	class 0	[Dierks and Krämer, 1988]
KiP1	180E-6	class 0	[Dierks and Krämer, 1988]
KiP2	2.8E-3	class 0	[Dierks and Krämer, 1988]
KcF	10.0	class 3	
KcR	10.0	class 3	
alpha	1.0	class 0	
beta	1.0	class 0	
gamma	1.0	class 0	
delta	1.0	class 0	
kinetic mechanism			Rapid Equilibrium Random Bi Bi
rate equation			[Sluse et al., 1991]
source for parameter estimation			See 7.12 -

## 4.5 AlaTA

Table 14: Kinetic parameters and their sources(AlaTA)

Parameter		class	notice
KmS1	2E-3	class 0	
KmS2	0.4E-3	class 0	
KmP1	32E-3	class 0	
KmP2	0.4E-3	class 0	
KiS1	8.7E-3	class 2	KiP2
KiP2	12E-3	class 0	
Keq	0.69	class 2	0.16, AspTA
KcF	337	class 0	at MW = 78000,
KcR	0.15	class 3	activity = 210 micromol/min/mg
kinetic mechanism			Ping-Pong Bi Bi
rate equation			[De Rosa et al., 1979]
source for parameter estimation			See 7.11 [De Rosa et al., 1979], Figure 3 with 5mM glutamate

## 4.6 AspTA

Table 15: Kinetic parameters and their sources(AspTA)

Parameter	class	notice
KmS1 0.9E-3	class 0	[Velick and Vavra, 1962, Table II]
KmS2 0.1E-3	class 0	[Velick and Vavra, 1962, Table II]
KmP1 0.04E-3	class 0	[Velick and Vavra, 1962, Table II]
KmP2 4E-3	class 0	[Velick and Vavra, 1962, Table II]
KiS1 2E-3	class 0	[Velick and Vavra, 1962, Table VII]
KiP2 8.3E-3	class 0	[Velick and Vavra, 1962, Table VII]
Keq 6.2	class 0	
KcF 300	class 0	
KcR 1000	class 0	from k4 and k10
kinetic mechanism		Ping-Pong Bi Bi [Velick and Vavra, 1962]
rate equation		See 7.11

## 4.7 CAC

Table 16: Kinetic parameters and their sources(CAC)

Parameter	class	notice
KmS1 0.6E-3	class 0	[Indiveri et al., 1994]
KmS2 9.4E-3	class 0	[Indiveri et al., 1994]
KmP1 43.4E-6	class 1	11.6E-6, the value of Car/Car reaction
KmP2 0.4E-3	class 1	1.2E-3, the value of Car/Car reaction
KiS1 8.7E-6	class 1	5.1E-6 [Indiveri et al., 1991b]
KiP2 250E-6	class 1	510E-6 [Indiveri et al., 1991b]
Keq 243.3	class 3	
KcF 1.22	class 2	
KcR 1.08	class 1	0.92, [Indiveri et al., 1991b]
kinetic mechanism		Ping-Pong Bi Bi [Indiveri et al., 1994]
rate equation		See 7.11
source for parameter estimation		[Indiveri et al., 1991b] Figure 4 with 13mM acetylcarnitine

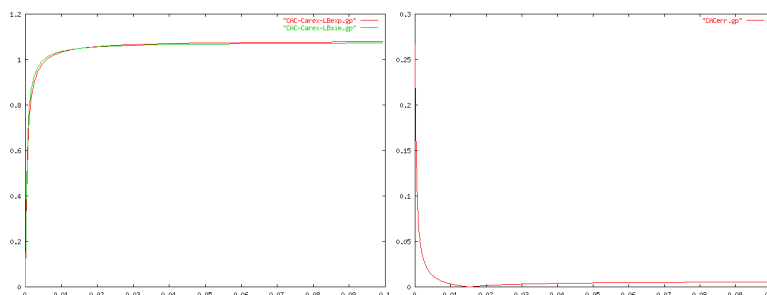


Figure 3: Comparison between experimental data and Computed data from estimated parameters(CAC)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.8 CIC

Table 17: Kinetic parameters and their sources(CIC)

Parameter		class	notice
KiS1	1.3E-4	class 2	11.2 mmol/min/g prot. $\times$ 30kDa 2.1, [Bisaccia et al., 1993, Table II]
KiS2	4.4E-4	class 2	
KiP1	3.3E-4	class 0	
KiP2	4.18E-5	class 0	
KcF	5.6	class 0	
KcR	3.5	class 1	
alpha	1.0	class 0	
beta	1.0	class 0	
gamma	1.0	class 0	
delta	1.0	class 0	
kinetic mechanism			Rapid Equilibrium Random Bi Bi [Bisaccia et al., 1993]
rate equation			See 7.12
source for parameter estimation			Figure 1(A) with 0.05mM citrate, (C) with 0.05mM malate [Bisaccia et al., 1993]

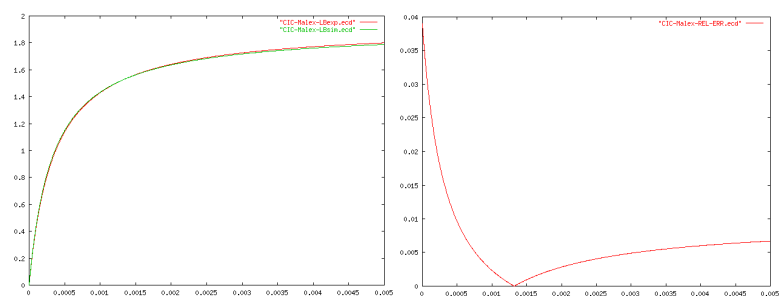


Figure 4: Comparison between experimental data and Computed data from estimated parameters(CIC)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.9 Complex I

Table 18: Kinetic parameters and their sources(Complex I)

Parameter	class	notice
KmS1	9.2E-6	class 0
KmS2	2.6E-4	class 0
KmP1	9.9E-6	class 2
KmP2	5.9E-5	class 2
KiS1	2.1E-8	class 0
KiP2	9.8E-8	class 2
Keq	407.9	class 3
KcF	498	class 0
KcR	229	class 2
kinetic mechanism		Ping-Pong Bi Bi
rate equation		[Fato et al., 1996]
source for parameter estimation		See 7.11
		[Fato et al., 1996]
		Figure 1(C) with 2.4 $\mu\text{M}$ reduced CoQ <sub>2</sub>

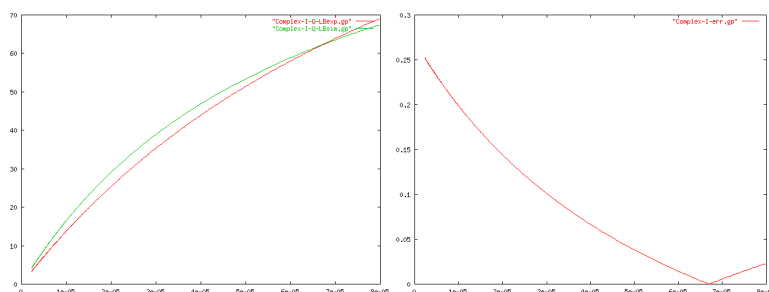


Figure 5: Comparison between experimental data and Computed data from estimated parameters(Complex I)

left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)

right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.10 Complex III

Table 19: Kinetic parameters and their sources(Complex III)

Parameter	class	notice	
KmA	2.8E-5	class 0	$K_5 \times KcF$
KmB	3.0E-6	class 0	$K_6 \times KcF$
Kb1	5.4E-6	class 2	$k_5/k_4$ $K_3 = K_4 \times Kb1$
Kb2	5.7E-6	class 2	$k_{10}/k_9$ , $K_1 = K_2 \times Kb2$
Kq1	2.8E-6	class 2	$k_7/k_6$ , $K_4 = Kq1/k_8$
Kq2	1.9E-6	class 2	$k_{12}/k_{11}$ , $K_2 = K_5 \times Kq2$
k8	622.1	class 2	
KcF	426.8	class 0	$1 / K_7$
kinetic mechanism			[Kubota et al., 1992, Scheme 3]
rate equation			See 7.3
source for parameter estimation			[Kubota et al., 1992] Figure 6 with $15 \mu\text{M}$ $Q_2H_2$

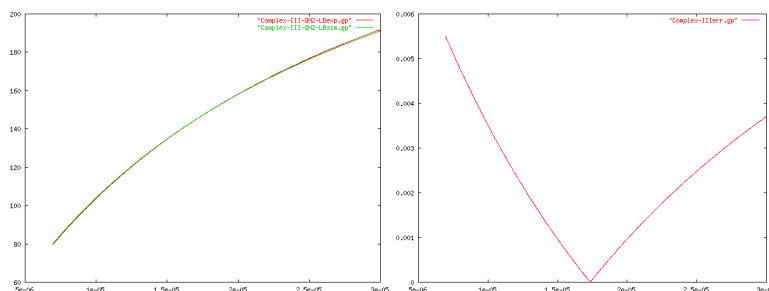


Figure 6: Comparison between experimental data and Computed data from estimated parameters(Complex III)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.11 Complex IV

Table 20: Kinetic parameters and their sources(Complex IV)

Parameter	class	notice	
Ks	110E-6	class 0	Value at pH = 7
KcF	93.5	class 0	Value at pH = 7, $\frac{d[\text{cyt2+}]}{dt} \times \frac{1}{4}$
kinetic mechanism			Michaelis Uni Uni
rate equation			[Malmström and Andréasson, 1985] See 7.7



## 4.12 Complex V

Table 21: Kinetic parameters and their sources(Complex V)

Parameter	class	notice
Kd	2.67E-7	class 3
Kp	9.02E-5	class 3
Kt	4.33E-5	class 3
KcF	14.5	class 0
Khx	1.3E-4	class 3
Khy	1.6E-4	class 3
klt_f	1.35E+8	class 3
klt_r	0.00018	class 3
ax	0.1	class 3
ay	0.6	class 3
beta	0.3	class 3
T	310	-
kinetic mechanism		see [Kholodenko, 1993]
rate equation		See 7.4
source for parameter estimation		[Matsuno-Yagi and Hatefi, 1985] Figure 2 with NADH respiration

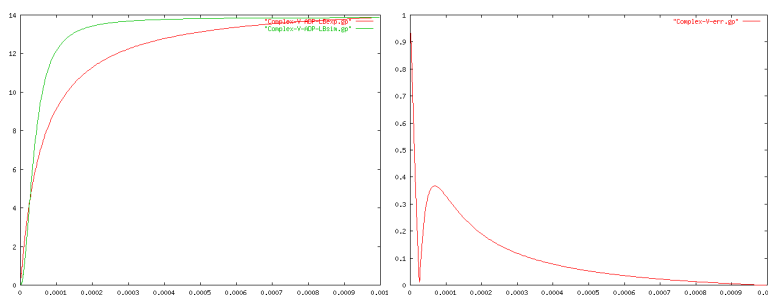


Figure 7: Comparison between experimental data and computed data from estimated parameters(Complex V)

left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)

right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

### 4.13 CPT I

Table 22: Kinetic parameters and their sources(CPT I)

Parameter		class	notice
KiS1	182E-6	class 0	[Ramsay et al., 1987]
KiS2	0.82E-6	class 0	
KiP1	6.7E-6	class 0	
KiP2	21E-6	class 0	
KcF	61.4	class 0	
KcR	32.8	class 0	
alpha	1.0	class 0	
beta	1.0	class 0	
gamma	1.0	class 0	
delta	1.0	class 0	
kinetic mechanism			
rate equation			[Ramsay et al., 1987] See 7.12

### 4.14 CPT II

Table 23: Kinetic parameters and their sources(CPT II)

Parameter		class	notice	
KmS1	6.3E-4	class 2	1.8 Unit/mg $\times$ 80kDa [Mann et al., 1995, Woeltje et al., 1987]	
KmS2	3.3E-4	class 2		
KmP1	950E-6	class 0		
KmP2	34E-6	class 0		
KiS1	2.4E-4	class 2		
KiS2	2.7E-4	class 2		
KiP1	41E-6	class 0		
KiP2	7E-6	class 0		
Keq	23540	class 3		
KcF	8.0	class 2		
KcR	2.4	class 0		
kinetic mechanism				Ordered Bi Bi
rate equation				[Mann et al., 1995] See 7.9
source for parameter estimation			[Mann et al., 1995] Figure 1 with $0\mu\text{M}$ SDZ	

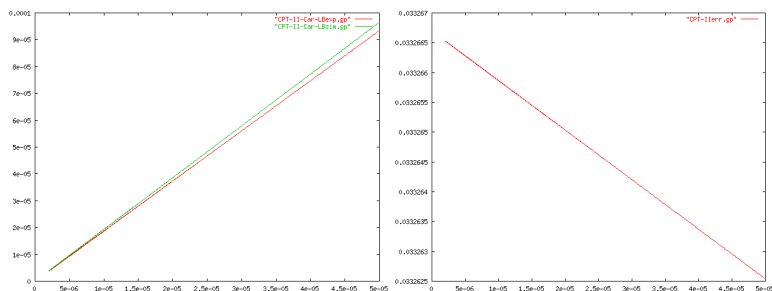


Figure 8: Comparison between experimental data and Computed data from estimated parameters(CPT II)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

#### 4.15 CS

Table 24: Kinetic parameters and their sources(CS)

Parameter	class	notice
k1	6.8E10	class 3
k_1	8.1E8	class 3
k2	3.0E10	class 3
k_2	7.2E8	class 3
k3	6.2E10	class 3
k_3	5.1E8	class 3
k4	1.2E10	class 3
k_4	4.0E8	class 3
k5	1.4E9	class 3
k_5	2.4E8	class 3
k6	4.1E10	class 3
k_6	1.1E8	class 3
k7	5E10	class 3
k_7	9.8E8	class 3
k8	5.3E10	class 3
k_8	7.7E8	class 3
kinetic mechanism		Random Bi Bi, [Shepherd and Garland, 1969], [Matsuoka and Srere, 1973], [Mukherjee and Srere, 1976]
source for parameter estimation		[Matsuoka and Srere, 1973]

## 4.16 DIC

Table 25: Kinetic parameters and their sources(DIC)

Parameter	class	notice
KiS1	0.20E-3	class 0
KiS2	0.72E-3	class 0
KiP1	9.0E-4	class 2
KiP2	7.6E-4	class 2
KcF	2.7	class 0
KcR	4.1	class 1
alpha	1.0	class 0
beta	1.0	class 0
gamma	1.0	class 0
delta	1.0	class 0
kinetic mechanism		Rapid Equilibrium Random Bi Bi
rate equation		[Indiveri et al., 1993]
source for parameter estimation		See 7.12
		Figure 5(A) with 0.05mM phosphate, (C) with 0.10mM malate [Indiveri et al., 1993]

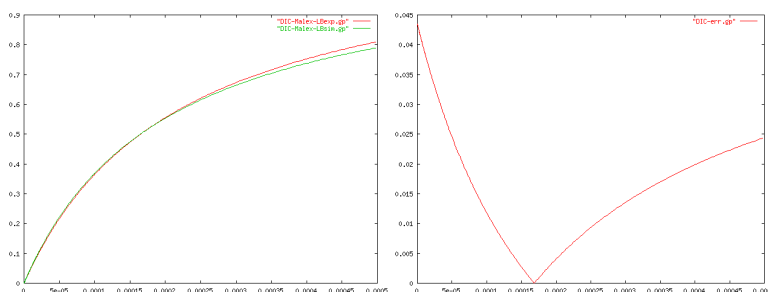


Figure 9: Comparison between experimental data and Computed data from estimated parameters(DIC)

left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)

right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.17 ECH

Table 26: Kinetic parameters and their sources(ECH)

Parameter	class	notice
Ks	16.9E-6	class 0
Kp	12.1E-6	class 0
KcF	8.9166667	class 0
KcR	2154.1667	class 0
kinetic mechanism		Uni Uni Reversible [Yang and Schulz, 1987]
rate equation		See 7.14

## 4.18 ETF-QO

Table 27: Kinetic parameters and their sources(ETF-QO)

Parameter		class	notice
KmS1	0.31E-6	class 0	
KmS2	0.39E-6	class 2	
KmP1	0.32E-6	class 0	
KmP2	4.2E-9	class 2	
KiS1	0.31E-6	class 0	
KiP2	0.3E-6	class 2	
Keq	0.66	class 0	
KcF	78	class 0	
KcR	101	class 2	
kinetic mechanism			Ping-Pong Bi Bi,
rate equation			[Beckmann and Frerman, 1985]
source for parameter estimation			See 7.11
			Figure 4 with 1.5 $\mu$ M
			ETF hydroquinone
			[Beckmann and Frerman, 1985]

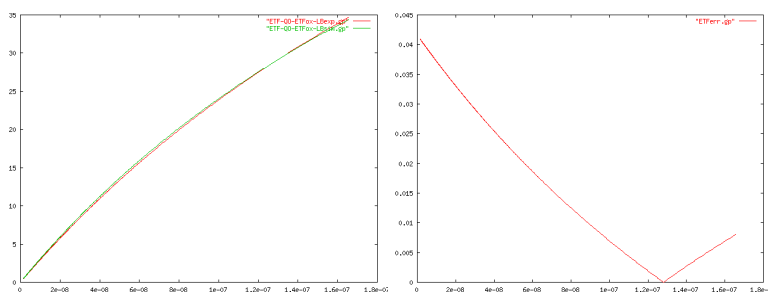


Figure 10: Comparison between experimental data and Computed data from estimated parameters(ETF:QO)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.19 FM

Table 28: Kinetic parameters and their sources(FM)

Parameter		class	notice
Ks	0.5E-5	class 0	[Boyer, 1970, Vol. V, chap. 19, Table V]
Kp	2.5E-5	class 0	
KcF	800	class 0	
KcR	900	class 0	
kinetic mechanism			Uni Uni Reversible
rate equation			See 7.14

## 4.20 HCD

Table 29: Kinetic parameters and their sources(HCD)

Parameter	class	notice
Ks 1.5E-6	class 0	
KcF 41.483333	class 0	
kinetic mechanism rate equation		Michaelis Uni Uni [Yang and Schulz, 1987] See 7.7

## 4.21 IDHa

Table 30: Kinetic parameters and their sources(IDHa)

Parameter	class	notice
KcF 105	class 0	28 U/mg $\times$ 224000 Da [Plaut et al., 1974, Ehrlich et al., 1981]
b 29.6	class 3	
c 0.00023	class 3	
d 7.8e-05	class 3	
e 0.00064	class 3	
f 0.00036	class 3	
kinetic mechanism rate equation source for parameter estimation		[Plaut et al., 1974] See 7.5 Figure 4 with 1.0mM ADP, [Plaut et al., 1974]

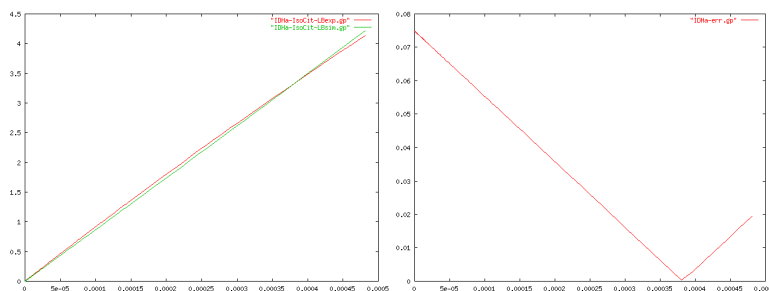


Figure 11: Comparison between experimental data and Computed data from estimated parameters(IDHa)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 4.22 IDHb

Table 31: Kinetic parameters and their sources(IDHb)

Parameter		class	notice	
phi0	5.1E-2	class 0	[Londesborough and Dalziel, 1970, Table 1]	
phi1	9.5E-8	class 0		
phi2	0.96E-6	class 0		
phi12	9E-8	class 0		
phir0	6.6E-2	class 0		
phir1	0.37E-6	class 0		
phir2	29E-6	class 0		
phir3	2.5E-4	class 0		
phir12	6E-12	class 0		
phir13	1.3E-10	class 0		
phir23	9.4E-8	class 0		
phir123	4.6E-14	class 0		
kinetic mechanism				See [Londesborough and Dalziel, 1970] See 7.6
rate equation				

## 4.23 MDH

Table 32: Kinetic parameters and their sources(MDH)

Parameter		class	notice
KmS1	72E-6	class 0	
KmS2	110E-6	class 0	
KmP1	1600E-6	class 0	
KmP2	170E-6	class 0	
KiS1	11E-6	class 0	
KiS2	100E-6	class 0	
KiP1	7100E-6	class 0	
KiP2	1900E-6	class 0	
KcF	0.390	class 0	
KcR	0.040	class 0	
kinetic mechanism			
rate equation			

## 4.24 NDK

Table 33: Kinetic parameters and their sources(NDK)

Parameter		class	notice
KmS1	0.31E-3	class 0	[Garces and Cleland, 1969]
KmS2	0.043E-3	class 0	[Garces and Cleland, 1969],UDP
KmP1	0.050E-3	class 0	[Garces and Cleland, 1969]
KmP2	0.25E-3	class 0	[Garces and Cleland, 1969],UTP
KiS1	0.21E-3	class 2	[Garces and Cleland, 1969]
KiP2	0.35E-3	class 2	[Garces and Cleland, 1969],UTP
Keq	1.28	class 0	[Garces and Cleland, 1969]
KcF	6883	class 0	MW = 70000 Da [Colomb et al., 1969]
KcR	5950	class 0	MW = 70000 Da [Colomb et al., 1969]
kinetic mechanism		Ping-Pong Bi Bi	
rate equation		[Garces and Cleland, 1969, Colomb et al., 1969]	
source for parameter estimation		See 7.11 [Colomb et al., 1969, Figure 4 with 0.18mM ATP]	

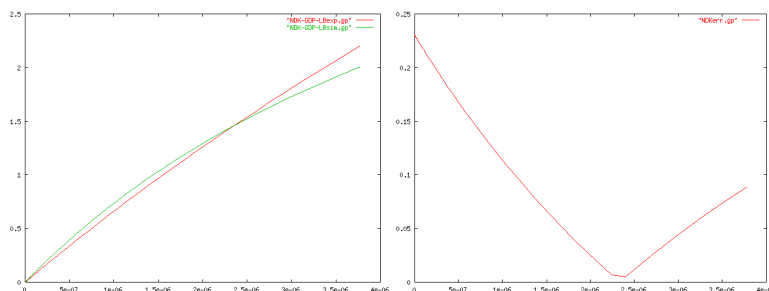


Figure 12: Comparison between experimental data and Computed data from estimated parameters(NDK)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error



## 4.25 OCT

Table 33: Kinetic parameters and their sources(OCT)

Parameter		class	notice
KmS1	1.1E-6	class 0	OCTa
	1.1E-6	class 0	OCTb, value for 16Oxoacyl-CoA
	1.3E-6	class 0	OCTc
	2.1E-6	class 0	OCTd
	3.2E-6	class 0	OCTe
	6.7E-6	class 0	OCTf
	12.4E-6	class 0	OCTg
KmS2	28.6E-6	class 0	
	28.6E-6	class 0	OCTb, value for 16Oxoacyl-CoA
	38.4E-6	class 0	OCTc
	35.7E-6	class 0	OCTd
	35.5E-6	class 0	OCTe
	18.9E-6	class 0	OCTf
	2.2E-6	class 0	OCTg
KmP1	7.2E-5	class 2	
KmP2	8.7E-5	class 2	
KiS1	1.1E-5	class 2	
KiP2	8.7E-5	class 2	
Keq	160.98	class 3	
KcF	137.86	class 0	$V_{max} \times 178000\text{Da}$
	137.86	class 0	OCTb, value for 16Oxoacyl-CoA
	253.52	class 0	OCTc
	272.94	class 0	OCTd
	277.38	class 0	OCTe
	264.07	class 0	OCTf
	80.244	class 0	OCTg
KcR	87.253	class 2	
	87.253	class 2	OCTb, value for 16Oxoacyl-CoA
	160.46	class 2	OCTc
	172.75	class 2	OCTd
	175.56	class 2	OCTe
	167.13	class 2	OCTf
	51.615	class 2	OCTg
kinetic mechanism			Ping-Pong Bi Bi, [Miyazawa et al., 1981]
rate equation			See 7.11
source for parameter estimation			Figure 5(B) with 200 $\mu\text{M}$ Acetyl-CoA, [Miyazawa et al., 1981]

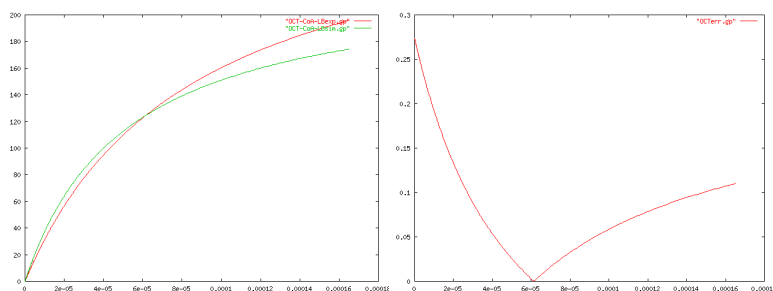


Figure 13: Comparison between experimental data and Computed data from estimated parameters(OCT)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

#### 4.26 OGC

Table 34: Kinetic parameters and their sources(OGC)

Parameter	class	notice
KiS1	0.3E-3	class 0
KiS2	0.7E-3	class 2
KiP1	1.4E-3	class 0
KiP2	0.17E-3	class 2
KcF	3.675	class 0
KcR	4.83	class 0
alpha	1.0	class 0
beta	1.0	class 0
gamma	1.0	class 0
delta	1.0	class 0
kinetic mechanism		Rapid Equilibrium Random Bi Bi, [Indiveri et al., 1991a]
rate equation		See 7.12
source for parameter estimation		Figure 2 with 20mM malate, [Indiveri et al., 1991a]

## 4.27 OGDC

Table 35: Kinetic parameters and their sources(OGDC)

Parameter	class	notice
KmA	0.22E-3	class 0
KmB	0.025E-3	class 0
KmC	0.050E-3	class 0
KmP	3E-4	class 2
KmR	6E-4	class 2
Kia	7.2E-4	class 2
Kib	7.4E-4	class 2
Kic	1E-4	class 2
Kip	1.1E-6	class 2
Kiq	81E-6	class 0
Kir	25E-6	class 0
KcF	177	class 2
kinetic mechanism		Multisite Ping-Pong
rate equation		[Cleland, 1973, Hamada et al., 1975]
source for parameter estimation		See 7.8
		Figure 1(A) with 0.010mM CoA,
		(B) with 0.20mM NAD,
		(C) with 0.10mM oxoglutarate
		[Hamada et al., 1975]

## 4.28 PC

Table 36: Kinetic parameters and their sources(PC)

Parameter	class	notice
KmA	0.11E-3	class 0
KmB	1.63E-3	class 0
KmC	0.37E-3	class 0
KmP	16E-3	class 0
KmQ	0.24E-3	class 0
KmR	0.051E-3	class 0
Keq	9.0	class 0
Kia	0.15E-3	class 0
Kib	1.6E-3	class 0
Kic	0.13E-3	class 0
Kip	7.9E-3	class 0
Kiq	0.19E-3	class 0
Kir	0.24E-3	class 0
KcF	200	class 0
KcR	20	class 0
kinetic mechanism		[Barden et al., 1972]
rate equation		See 7.10

## 4.29 PDC

Table 37: Kinetic parameters and their sources(PDC)

Parameter	class	notice
KmA	25E-6	class 0
KmB	13E-6	class 0
KmC	50E-6	class 0
KmP	5.9E-7	class 2
KmR	6.9E-7	class 2
Kia	5.5E-4	class 2
Kib	3.0E-4	class 2
Kic	1.8E-4	class 2
Kip	6.0E-5	class 2
Kiq	35E-6	class 0
Kir	36E-6	class 0
KcF	856	class 1
kinetic mechanism		Multisite Ping-Pong, [Cleland, 1973, Hamada et al., 1975]
rate equation		See 7.8
source for parameter estimation		Figure 2(A) with 0.015mM CoA, (B) with 0.050mM NAD, (C) with 0.050mM pyruvate [Hamada et al., 1975]

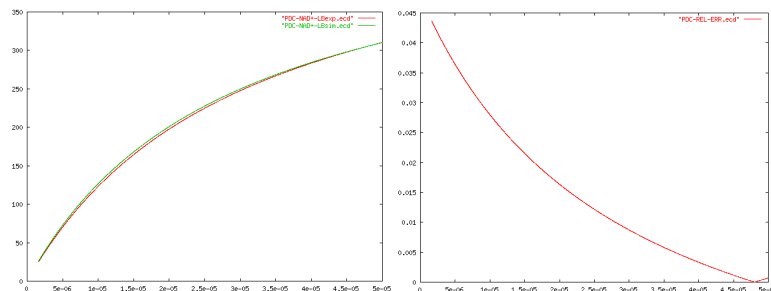


Figure 14: Comparison between experimental data and Computed data from estimated parameters(PDC)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

### 4.30 PiC

Table 38: Kinetic parameters and their sources(PiC)

Parameter		class	notice
KiS1	0.87	class 2	
KiS2	1.86E-8	class 2	
KiP1	32.84E-9	class 0	Fig. 4, [Stappen and Krämer, 1994]
KiP2	11.12E-3	class 0	Fig. 4, [Stappen and Krämer, 1994]
KcF	37.9	class 0	Fig. 4, [Stappen and Krämer, 1994], 34kDa
KcR	37.0	class 0	Fig. 4, [Stappen and Krämer, 1994], 34kDa
alpha	1.0	class 0	
beta	1.0	class 0	
gamma	1.0	class 0	
delta	1.0	class 0	
kinetic mechanism			Rapid Equilibrium Random Bi Bi, [Stappen and Krämer, 1994]
rate equation			See 7.12
source for parameter estimation			Figure 4(A) with pH5.85, (B) with 4mM phosphate]stappen94

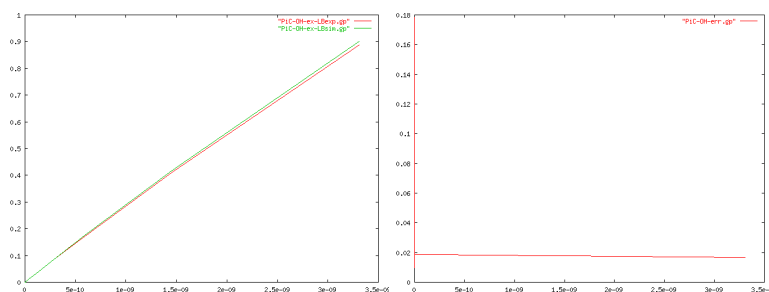


Figure 15: Comparison between experimental data and Computed data from estimated parameters(PiC)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

### 4.31 PYC

Table 39: Kinetic parameters and their sources(PYC)

Parameter		class	notice
KiS1	6.1E-4	class 2	
KiS2	5.9E-4	class 2	
KiP1	2.6E-4	class 2	
KiP2	4.1E-4	class 2	
KcF	0.84	class 1	
KcR	0.78	class 1	
alpha	1.0	class 0	
beta	1.0	class 0	
gamma	1.0	class 0	
delta	1.0	class 0	
kinetic mechanism			Rapid Equilibrium Random Bi Bi, (sequential) [Nalecz, 1994]
rate equation			See 7.12
source for parameter estimation			[Capuano et al., 1990, Figure 3]

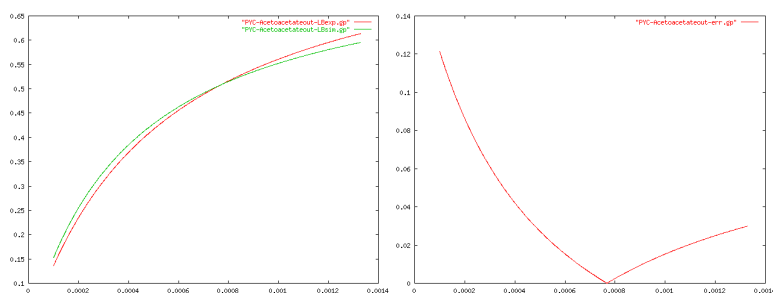


Figure 16: Comparison between experimental data and Computed data from estimated parameters(PYC)

- left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)
- right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

### 4.32 SCS

Table 40: Kinetic parameters and their sources(SCS)

Parameter	class	notice
KmA	5E-6	class 0
KmB	3.5E-5	class 0
KmC	4.5E-4	class 0
KmP	6E-4	class 0
KmQ	7.5E-6	class 0
KmC2	4.5E-4	class 0
KmP2	6E-4	class 0
Keq	8.375	class 0
Kia	4E-4	class 0
Kib	2E-5	class 0
Kic	3E-5	class 0
Kip	7E-2	class 0
Kiq	5E-6	class 0
Kir	6.7E-6	class 0
Kc1	100	class 0
Kc2	100	class 3
kinetic mechanism		See [Cha and Parks Jr., 1964]
rate equation		See 7.13

### 4.33 SDH

Table 41: Kinetic parameters and their sources(SDH)

Parameter	class	notice
KmS1	30E-6	class 0
KmS2	69E-6	class 0
KmP1	0.3E-6	class 0
KmP2	1.5E-6	class 0
KiS1	4.1E-6	class 2
KiP2	5.6E-6	class 2
Keq	0.037	class 0
KcF	69.3	class 0
KcR	1.73	class 0
kinetic mechanism		Ping-Pong Bi Bi [Grivennikova et al., 1993]
rate equation		See 7.11
source for parameter estimation		[Grivennikova et al., 1993, Figure 2(B)]

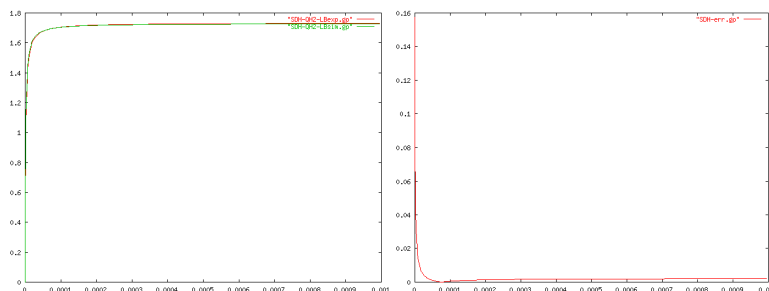


Figure 17: Comparison between experimental data and Computed data from estimated parameters(SDH)

left: Comparison between experimental data and computed ones  
 abscissa = reaction rate ( $\text{sec}^{-1}$ )  
 ordinate = substrate concentration (M)

right: Percent error between experimental data and computed ones  
 abscissa = substrate concentration (M)  
 ordinate = percent error

## 5 Initial condition

### 5.1 The respiratory chain

Table 42: Initial condition of enzymes (respiratory chain)

name	localization	number of molecules
Complex-I	MT-IM	1000
Complex-III	MT-IM	3000
Complex-IV	MT-IM	7000
Complex-V	MT-IM	900

Table 43: Initial condition of metabolites (respiratory chain)

name	localization	concentration
Q	MT-IMS	0.26E-3 M
QH2	MT-IMS	0.028E-3 M
Cyt-c3+	MT-IMS	3E-6 M
Cyt-c2+	MT-IMS	0.11E-3 M
H+	MT-IMS	1E-6 M (fix)
H+	MATRIX	1E-8 M (fix)

### 5.2 The TCA cycle

Table 44: Initial condition of enzymes (TCA cycle)

name	localization	number of molecules
CS	MATRIX	100
ACO	MATRIX	100
IDHa	MATRIX	100
IDHb	MATRIX	100
OGDC	MATRIX	100
SCS	MATRIX	100
SDH	MT-IM	100
FM	MATRIX	100
MDH	MATRIX	100
AlaTA	MATRIX	100
AspTA	MATRIX	100
NDK	MATRIX	100
PDC	MATRIX	100
PC	MATRIX	100



Table 45: Initial condition of metabolites (TCA cycle)

name	localization	concentration
Cit	MT-IMS	0.42E-3 M (fix)
Cit	MATRIX	0.42E-3 M
IsoCit	MATRIX	0.42E-3 M
OG	MT-IMS	0.021E-3 M (fix)
OG	MATRIX	0.021E-3 M
SCoA	MATRIX	76168
Suc	MATRIX	2.95E-3 M
Fum	MATRIX	0.065E-3 M
Mal	MT-IMS	0.50E-3 M (fix)
Mal	MATRIX	0.50E-3 M
OXA	MATRIX	0.004E-3 M
Asp	MATRIX	1.14E-3 M
Asp	MT-IMS	1.14E-3 M (fix)
Glu	MATRIX	3.03E-3 M
Glu	MT-IMS	3.03E-3 M (fix)
Ala	MATRIX	3.44E-3 M
Pyr	MT-IMS	0.1025E-3 M (fix)
Pyr	MATRIX	0.1025E-3 M (fix)
CoA	MT-IMS	700 (fix)
CoA	MATRIX	70435
Acetyl-CoA	MATRIX	0.03E-3 M
NADH	MATRIX	0.072E-3 M
NAD+	MATRIX	0.170E-3 M
NADPH	MATRIX	0.072E-3 M
NADP+	MATRIX	0.170E-3 M
CO2	MATRIX	1.63E-3 M

Table 46: Initial condition of metabolites

name	localization	concentration
ATP	MT-IMS	4.5E-3 M (fix)
ATP	MATRIX	4.5E-3 M
ADP	MT-IMS	0.45E-3 M (fix)
ADP	MATRIX	0.45E-3 M (fix)
GTP	MATRIX	4.5E-3 M
GDP	MATRIX	0.45E-3 M
Pi	MT-IMS	4E-3 M (fix)
Pi	MATRIX	4E-3 M

### 5.3 Fatty acid $\beta$ oxidation

Table 47: Initial condition of enzymes (fatty acid  $\beta$  oxidation)

name	localization	number of molecules
CPT-I	MT-OM	100
CAC	MT-IM	100
ACD	MT-IM	100
ECH	MT-IM	100
HCD	MT-IM	100
OCT	MT-IM	100
ETF-QO	MT-IM	100

Table 48: Initial condition of metabolites (fatty acid  $\beta$  oxidation 1)

name	localization	concentration
Car	MT-IMS	0.2E-3 M (fix)
Car	MATRIX	0.95E-3 M
PalCar	MT-IMS	0.6E-3 M (fix)
PalCar	MATRIX	0.012E-3 M
16Acyl-CoA	MT-IMS	0.039E-3 M (fix)
ETFred	MATRIX	0.31E-6 M
ETFox	MATRIX	0.32E-6 M

Table 49: Initial condition of metabolites (fatty acid  $\beta$  oxidation 2)

name	localization	concentration
16Acyl-CoA	MATRIX	0.039E-3 M
16Enoyl-CoA	MATRIX	0.017E-3 M
16Hydroxyacyl-CoA	MATRIX	0.012E-3 M
16Oxoacyl-CoA	MATRIX	0.0011E-3 M
14Acyl-CoA	MATRIX	0.039E-3 M
14Enoyl-CoA	MATRIX	0.017E-3 M
14Hydroxyacyl-CoA	MATRIX	0.012E-3 M
14Oxoacyl-CoA	MATRIX	0.0011E-3 M
12Acyl-CoA	MATRIX	0.087E-3 M
12Enoyl-CoA	MATRIX	0.017E-3 M
12Hydroxyacyl-CoA	MATRIX	0.012E-3 M
12Oxoacyl-CoA	MATRIX	0.0013E-3 M
10Acyl-CoA	MATRIX	0.087E-3 M
10Enoyl-CoA	MATRIX	0.017E-3 M
10Hydroxyacyl-CoA	MATRIX	0.012E-3 M
10Oxoacyl-CoA	MATRIX	0.0021E-3 M
8Acyl-CoA	MATRIX	0.087E-3 M
8Enoyl-CoA	MATRIX	0.017E-3 M
8Hydroxyacyl-CoA	MATRIX	0.012E-3 M
8Oxoacyl-CoA	MATRIX	0.0032E-3 M
6Acyl-CoA	MATRIX	0.087E-3 M
6Enoyl-CoA	MATRIX	0.017E-3 M
6Hydroxyacyl-CoA	MATRIX	0.012E-3 M
6Oxoacyl-CoA	MATRIX	0.0067E-3 M
4Acyl-CoA	MATRIX	0.087E-3 M
4Enoyl-CoA	MATRIX	0.017E-3 M
4Hydroxyacyl-CoA	MATRIX	0.012E-3 M
Acetoacetyl-CoA	MATRIX	0.0124E-3 M

## 5.4 The inner-membrane transport

Table 50: Initial condition of enzymes (inner-membrane transport)

name	localization	number of molecules
AAC	MT-IM	1000
AGC	MT-IM	1000
PiC	MT-IM	1000
PYC	MT-IM	1000
OGC	MT-IM	1000
DIC	MT-IM	1000
CIC	MT-IM	1000

## 6 Steady-state condition

This metabolic model reached in a steady-state around  $t=93000\text{sec}$  with the initial condition. Number of molecules at the steady-state are shown below. Obtaining a steady-state, this system clears requirements for Metabolic Control Analysis[Fell, 1992].

### 6.1 The respiratory chain

Table 51: Steady-state concentration of metabolites (respiratory chain)

name	localization	number of molecules
Q	MT-IMS	77547
QH2	MT-IMS	500
Cyt-c3+	MT-IMS	29624
Cyt-c2+	MT-IMS	999
H+	MT-IMS	3
H+	MATRIX	3

Table 52: Steady-state concentration of metabolites

name	localization	number of molecules
ATP	MT-IMS	13550(fix)
ATP	MATRIX	180
ADP	MT-IMS	1355(fix)
ADP	MATRIX	121948(fix)
GTP	MATRIX	2579
GDP	MATRIX	1338852
Pi	MT-IMS	12044(fix)
Pi	MATRIX	2507395

## 6.2 The TCA cycle and the inner-membrane transport

Table 53: Steady-state concentration of metabolites (TCA cycle)

name	localization	number of molecules
Cit	MT-IMS	1265 (fix)
Cit	MATRIX	583455
IsoCit	MATRIX	74758
OG	MT-IMS	63 (fix)
OG	MATRIX	424
SCoA	MATRIX	32
Suc	MATRIX	1133
Fum	MATRIX	231567
Mal	MT-IMS	1506 (fix)
Mal	MATRIX	1028383
OXA	MATRIX	302
Asp	MATRIX	244090
Asp	MT-IMS	3433 (fix)
Glu	MATRIX	801482
Glu	MT-IMS	9124 (fix)
Ala	MATRIX	1016709
Pyr	MT-IMS	27777 (fix)
Pyr	MATRIX	309(fix)
CoA	MT-IMS	700 (fix)
CoA	MATRIX	286
Acetyl-CoA	MATRIX	104498
NADH	MATRIX	3672
NAD+	MATRIX	61909
NADPH	MATRIX	7508
NADP+	MATRIX	58073
CO2	MATRIX	42631671

## 6.3 Fatty acid $\beta$ oxidation

Table 54: Steady-state concentration of metabolites (fatty acid  $\beta$  oxidation 1)

name	localization	number of molecules
Car	MT-IMS	602 (fix)
Car	MATRIX	47418
PalCar	MT-IMS	1807 (fix)
PalCar	MATRIX	213280
16Acyl-CoA	MT-IMS	117 (fix)
ETFred	MATRIX	89
ETFox	MATRIX	82

Table 55: Steady-state concentration of metabolites (fatty acid  $\beta$  oxidation 2)

name	localization	number of molecules
16Acyl-CoA	MATRIX	331
16Enoyl-CoA	MATRIX	698
16Hydroxyacyl-CoA	MATRIX	3
16Oxoacyl-CoA	MATRIX	769
14Acyl-CoA	MATRIX	331
14Enoyl-CoA	MATRIX	699
14Hydroxyacyl-CoA	MATRIX	3
14Oxoacyl-CoA	MATRIX	771
12Acyl-CoA	MATRIX	330
12Enoyl-CoA	MATRIX	700
12Hydroxyacyl-CoA	MATRIX	2
12Oxoacyl-CoA	MATRIX	763
10Acyl-CoA	MATRIX	331
10Enoyl-CoA	MATRIX	700
10Hydroxyacyl-CoA	MATRIX	2
10Oxoacyl-CoA	MATRIX	762
8Acyl-CoA	MATRIX	332
8Enoyl-CoA	MATRIX	701
8Hydroxyacyl-CoA	MATRIX	2
8Oxoacyl-CoA	MATRIX	763
6Acyl-CoA	MATRIX	332
6Enoyl-CoA	MATRIX	701
6Hydroxyacyl-CoA	MATRIX	3
6Oxoacyl-CoA	MATRIX	764
4Acyl-CoA	MATRIX	331
4Enoyl-CoA	MATRIX	702
4Hydroxyacyl-CoA	MATRIX	2
Acetoacetyl-CoA	MATRIX	239686

## 7 Rate equations

### 7.1 AAC

$$v = \frac{\frac{k_{\rightarrow}^D(\Delta\psi)[E_{total}][ADP_{out}]}{1 + \frac{k_{\rightarrow}^D(\Delta\psi)}{k_{\leftarrow}^D(\Delta\psi)} \left(1 + \frac{K^{D'}}{[ADP_{in}]}\right)}}{\frac{K^{D'}}{1 + \frac{k_{\rightarrow}^D(\Delta\psi)}{k_{\leftarrow}^D(\Delta\psi)} \left(1 + \frac{K^{D'}}{[ADP_{in}]}\right)} + [ADP_{out}]}$$

$$\begin{aligned} k_{\rightarrow}^D(\Delta\psi) &= k_{0\rightarrow}^D e^{\phi C_f} \cdot \text{normalize} \\ k_{\leftarrow}^D(\Delta\psi) &= k_{0\leftarrow}^D e^{\phi C_r} \cdot \text{normalize} \end{aligned}$$

$$\phi = \frac{RT}{F} \ln \frac{[H_{IMS}]}{[H_{MAT}]}$$

### 7.2 CB Ordered Bi Bi

$$\begin{aligned} v &= \frac{\left(\frac{K_c F [S1][S2]}{K_{iS1} K_{mS2}} - \frac{K_c R [P1][P2]}{K_{mP1} K_{iP2}}\right) [E]}{\text{denom}} \\ \text{denom} &= 1 + \frac{[S1]}{K_{iS1}} + \frac{K_{mS1} [S2]}{K_{iS1} K_{mS2}} + \frac{K_{mP2} [P1]}{K_{mP1} K_{iP2}} + \frac{[P2]}{K_{iP2}} + \frac{[S1][S2]}{K_{iS1} K_{mS2}} \\ &+ \frac{K_{mP2} [S1][P1]}{K_{iS1} K_{mP1} K_{iP2}} + \frac{K_{mS1} [S2][P2]}{K_{iS1} K_{mS2} K_{iP2}} + \frac{[P1][P2]}{K_{mP1} K_{iP2}} + \frac{[S1][S2][P1]}{K_{iS1} K_{mS2} K_{iP1}} + \frac{[S2][P1][P2]}{K_{iS2} K_{mP1} K_{iP2}} \end{aligned}$$

### 7.3 Complex III

$$v = \frac{K_{cF}[E_t][A][B]}{\text{denom}}$$

$$\text{denom} = \left( K_{mA}K_{q2}K_{b2} + K_{mA}K_{q2}[B] + \frac{K_{cF}}{k_8}K_{q1}[A]K_{b1} + \frac{K_{cF}}{k_8}K_{q1}[A][B] \right) [Q]$$

$$+ K_{mA}[B] + K_{mB}[A] + [A][B]$$

### 7.4 Complex V

$$v = \frac{K_{cF}[E] \left\{ \frac{[ADP][Pi]}{K_d K_p} k_{ltf} e^{-3(\beta - a_x)\phi} \left( \frac{[H_{IMS}^+]}{K_{hx e^{ax}\phi}} \right)^3 - \frac{[ATP]}{K_t} K_{eq} k_{ltr} e^{3(1 - \beta - a_y)\phi} \left( \frac{[H_{MAT}^+]}{K_{hy e^{-ay}\phi}} \right)^3 \right\}}{\left( 1 + \frac{[H_{IMS}^+]}{K_{hx e^{ax}\phi}} + \frac{[H_{MAT}^+]}{K_{hy e^{-ay}\phi}} \right)^3 \left( 3 + \frac{[ADP][Pi]}{K_d K_p} + \frac{[ATP]}{K_t} \right)}$$

$$\text{where } \phi = \ln \frac{[H_{IMS}^+]}{[H_{MAT}^+]}$$

### 7.5 IDHa

$$v = \frac{k_{cat}[E]([IsoCit]^2 + b[ADP][IsoCit])}{[IsoCit]^2 + c[IsoCit] + d[ADP] + e[ADP][IsoCit] + f}$$

### 7.6 IDHb

$$v = \frac{[E][NADP][IsoCit]}{\text{denom1}} - \frac{[E][NADPH][OG][CO_2]}{\text{denom2}}$$

$$\text{denom1} = \phi_0[NADP][IsoCit] + \phi_1[IsoCit] + \phi_2[NADP] + \phi_{12}$$

$$\text{denom2} = \phi'_0[NADPH][OG][CO_2] + \phi'_1[OG][CO_2] + \phi'_2[NADPH][CO_2] + \phi'_3[NADPH][OG]$$

$$+ \phi'_{12}[CO_2] + \phi'_{13}[OG] + \phi'_{23}[NADPH] + \phi'_{123}$$

### 7.7 Michaelis Uni Uni

$$v = \frac{K_{cF}[E][S]}{K_s + [S]}$$

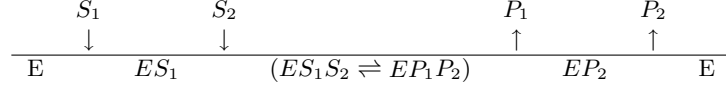
### 7.8 Multisite Ping-Pong

$$v = \frac{k_{cat}[E_{total}][A][B][C]}{\text{denom}}$$

$$\text{denom} = K_{mC}[A][B] + K_{mB}[A][C] + K_{mA}[B][C] + [A][B][C]$$

$$+ \frac{K_{mA}K_{mP}K_{ib}K_{ic}[Q][R]}{K_{mR}K_{ip}K_{iq}} + \frac{K_{mC}[A][B][R]}{K_{ir}} + \frac{K_{mB}[A][C][Q]}{K_{iq}} + \frac{K_{mA}K_{mP}K_{ib}K_{ic}[A][Q][R]}{K_{mR}K_{ip}K_{ia}K_{iq}}$$

## 7.9 Ordered Bi Bi



$$v = \frac{K_{cF}K_{cR}[E]([S_1][S_2] - \frac{[P_1][P_2]}{K_{eq}})}{\text{denom}}$$

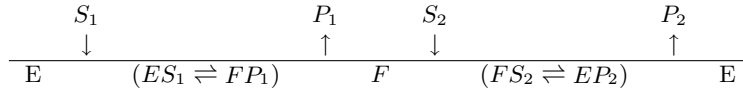
$$\begin{aligned}
 \text{denom} &= K_{cR}K_{iS_1}K_{mS_2} + K_{cR}K_{mS_2}[S_1] + K_{cR}K_{mS_1}[S_2] + \frac{K_{cF}K_{mP_2}[P_1]}{K_{eq}} + \frac{K_{cF}K_{mP_1}[P_2]}{K_{eq}} \\
 &+ K_{cR}[S_1][S_2] + \frac{K_{cF}K_{mP_2}[S_1][P_1]}{K_{eq}K_{iS_1}} + \frac{K_{cF}[P_1][P_2]}{K_{eq}} + \frac{K_{cR}K_{mS_1}[S_2][P_2]}{K_{iQ}} + \frac{K_{cR}[S_1][S_2][P_1]}{K_{iP_1}} \\
 &+ \frac{K_{cF}[S_2][P_1][P_2]}{K_{iS_2}K_{eq}}
 \end{aligned}$$

## 7.10 PC

$$v = \frac{V_1V_2[A][B][C] - \frac{V_1V_2[P][Q][R]}{K_{eq}}}{\text{denom}}$$

$$\begin{aligned}
 \text{denom} &= K_{iA}K_{mB}V_2[C] + K_{mC}V_2[A][B] + K_{mA}V_2[B][C] + K_{mB}V_2[B][C] + K_{mB}V_2[A][C] + V_2[A][B][C] \\
 &+ \frac{K_{iP}K_{mQ}V_1[R]}{K_{eq}} + \frac{K_{mQ}V_1[P][R]}{K_{eq}} + \frac{K_{mP}V_1[Q][R]}{K_{eq}} + \frac{K_{mR}V_1[P][Q]}{K_{eq}} + \frac{V_1[P][Q][R]}{K_{eq}} \\
 &+ \frac{K_{iA}K_{mB}V_2[C][P]}{K_{iP}} + \frac{K_{iA}K_{mB}V_2[C][Q]}{K_{iP}} + \frac{K_{iQ}K_{mP}V_1[B][R]}{K_{iB}K_{eq}} + \frac{K_{iQ}K_{mP}V_1[A][R]}{K_{iA}K_{eq}} \\
 &+ \frac{K_{iA}V_2[A][B][R]}{K_{iR}} + \frac{K_{mR}V_1[C][P][Q]}{K_{iC}K_{eq}} + \frac{K_{mA}V_2[B][C][Q]}{K_{iQ}} + \frac{K_{mA}V_2[B][C][P]}{K_{iP}} \\
 &+ \frac{K_{mP}V_1[B][Q][R]}{K_{iB}K_{eq}} + \frac{K_{mQ}V_1[B][P][R]}{K_{iB}K_{eq}}
 \end{aligned}$$

## 7.11 Ping-Pong Bi Bi



$$v = \frac{K_{cF}K_{cR}[E]([S_1][S_2] - \frac{[P_1][P_2]}{K_{eq}})}{\text{denom}}$$

$$\begin{aligned}
 \text{denom} &= K_{cR}K_{mS_2}[S_1] + K_{cR}K_{mS_1}[S_2] + \frac{K_{cF}K_{mP_2}[P_1]}{K_{eq}} + \frac{K_{cF}K_{mP_1}[P_2]}{K_{eq}} + K_{cR}[S_1][S_2] \\
 &+ \frac{K_{cF}K_{mP_2}[S_1][P_1]}{K_{eq}K_{iS_1}} + \frac{K_{cF}[P_1][P_2]}{K_{eq}} + \frac{K_{cR}K_{mS_1}[S_2][P_2]}{K_{iQ}}
 \end{aligned}$$

## 7.12 Rapid Equilibrium Random Bi Bi

$$v = \frac{\frac{[A][B]}{\alpha K_{iA} K_{iB}} k_{cat}^f [E]_{total} - \frac{[P][Q]}{\beta K_{iP} K_{iQ}} k_{cat}^r [E]_{total}}{1 + \frac{[A]}{K_{iA}} + \frac{[B]}{K_{iB}} + \frac{[P]}{K_{iP}} + \frac{[Q]}{K_{iQ}} + \frac{[A][B]}{\alpha K_{iA} K_{iB}} + \frac{[P][Q]}{\beta K_{iP} K_{iQ}} + \frac{[B][Q]}{\gamma K_{iB} K_{iQ}} + \frac{[A][P]}{\delta K_{iA} K_{iP}}}$$

## 7.13 SCS

$$v = \frac{\left( [A][B][C] - \frac{[P][Q][R]}{K_{eq}} \right) \left\{ V_1 + V_2 \left( \frac{K_{mC}[P]}{K_{mC2} K_{ip}} + \frac{[C]}{K_{mC2}} \right) \right\}}{\text{denom}}$$

$$\begin{aligned} \text{denom} &= K_{ia} K_{mB} [C] + K_{mB} [A][C] + K_{mA} [B][C] + K_{mC} [A][B] + [A][B][C] \\ &+ \frac{[A][B][C]^2}{K_{mC2}} + \frac{K_{ia} K_{mB} K_{mC} [P]}{K_{ip}} + \frac{K_{ia} K_{mB} K_{mC} [P][Q]}{K_{ip} K_{iq}} + \frac{K_{ia} K_{mB} K_{mC} [P][R]}{K_{ip} K_{ir}} \\ &+ \frac{K_{ia} K_{mB} K_{ic} [Q][R]}{K_{mQ} K_{ir}} + \frac{K_{ia} K_{mB} K_{mC} [P][Q][R]}{K_{ip} K_{mQ} K_{ir}} + \frac{K_{ia} K_{mB} K_{mC} [P]^2 [Q][R]}{K_{ip} K_{mP2} K_{mQ} K_{ir}} \\ &+ \frac{K_{ia} K_{mB} [C][Q]}{K_{iq}} + \frac{K_{ia} K_{mB} [C][R]}{K_{ir}} + \frac{K_{ia} K_{mB} [C][Q][R]}{K_{mQ} K_{ir}} + \frac{K_{ia} K_{mB} [C][P][Q][R]}{K_{mP2} K_{mQ} K_{ir}} \\ &+ \frac{K_{mB} K_{mC} [A][P]}{K_{ip}} + \frac{K_{mA} K_{mC} [B][P]}{K_{ip}} + \frac{K_{mC} [A][B][P]}{K_{ip}} + \frac{K_{mC} [A][B][C][P]}{K_{mC2} K_{ip}} \\ &+ \frac{K_{mA} [B][C][Q]}{K_{iq}} + \frac{K_{mB} [A][C][R]}{K_{ir}} + \frac{K_{mA} K_{mC} [B][P][Q]}{K_{ip} K_{iq}} + \frac{K_{mB} K_{mC} [A][P][R]}{K_{ip} K_{ir}} \end{aligned}$$

## 7.14 Uni Uni Reversible

$$v = \frac{(K_{cF} K_p [S] - K_{cR} K_s [P]) [E]}{K_s [P] + K_p [S] + K_s K_p}$$



## 8 MeSH term and literature search

The mitochondrial model was built through comprehensive literature search. Here we show the tendency of MeSH terms embedded in the articles that were crucial for determination of the rate equations.

Table 56: The MeSH term tendency of the articles on the reaction mechanism of each enzyme

Literature	Kinetics	Models	Mathematics	enzyme name	substrate name
[Barden et al., 1972]	+	+, Chemical	+	-	+
[Beckmann and Frerman, 1985]	+	-	-	+	-
[Crow et al., 1983]	+	-	+	+	+
[Davisson and Schulz, 1985]	+	+, Biological	-	+	+
[De Rosa et al., 1979]	-	-	-	+	-
[Dierks and Krämer, 1988]	+	-	-	+	-
[Fato et al., 1996]	+	-	-	+	+
[Grivennikova et al., 1993]	-	-	-	+	-
[Guarriero-Bobyleva et al., 1978]	+	-	-	+	+
[Hamada et al., 1975]	+	-	-	+	+
[Indiveri et al., 1991b]	+	-	-	+	+
[Indiveri et al., 1991a]	+	-	-	+	+
[Indiveri et al., 1994]	+	-	-	+	+
[Kholodenko, 1993]	+	+	-	+	+
[Krämer and Klingenberg, 1982]	+	-	-	+	+
[Kubota et al., 1992]	+	-	-	+	+
[Malmström and Andréasson, 1985]	+	-	-	+	+
[Mann et al., 1995]	+	-	-	+	+
[Matsuoka and Srere, 1973]	+	-	+	+	+
[McKean et al., 1979]	+	-	-	-	+
[Miyazawa et al., 1981]	+	-	-	+	-
[Mukherjee and Srere, 1976]	-	-	-	-	+
[Plaut et al., 1974]	+	+, Chemical	+	+	+
[Ramsay et al., 1987]	+	-	-	+	+
[Sluse et al., 1991]	+	-	-	+	+
[Stappen and Krämer, 1994]	+	-	-	+	-
[Yang and Schulz, 1987]	+	+, Theoretical	+	+	+
Frequency	24/27	5/27	5/27	24/27	21/27

## References

- [Barden et al., 1972] Barden, R. E., Fung, C. H., Utter, M. F., and Scrutton, M. C. (1972). Pyruvate carboxylase from chicken liver. *J. Biol. Chem.*, 247(4):1323–33.
- [Beckmann and Frerman, 1985] Beckmann, J. D. and Frerman, F. E. (1985). Reaction of electron-transfer flavoprotein with electron-transfer flavoprotein-ubiquinone oxidoreductase. *Biochemistry*, 24(15):3922–5.
- [Bisaccia et al., 1993] Bisaccia, F., De Palma, A., Dierks, T., Krämer, R., and Palmieri, F. (1993). Reaction mechanism of the reconstituted tricarboxylate carrier from rat liver mitochondria. *Biochim. Biophys. Acta*, 1142(1-2):139–45.
- [Boyer, 1970] Boyer, P. D., editor (1970). *The Enzymes*. Academic Press, New York, London.
- [Capuano et al., 1990] Capuano, F., Di Paola, M., Azzi, A., and Papa, S. (1990). The monocarboxylate carrier from rat liver mitochondria: purification and kinetic characterization in a reconstituted system. *FEBS lett.*, 261(1):39–42.
- [Cha and Parks Jr., 1964] Cha, S. and Parks Jr., R. E. (1964). Succinic thiokinase II. kinetic studies: initial velocity, product inhibition, and effect of arsenate. *J. Biol. Chem.*, 239(6):1968–77.
- [Cleland, 1973] Cleland, W. W. (1973). Derivation of rate equations of multisite ping-pong mechanisms with ping-pong reactions at one or more sites. *J. Biol. Chem.*, 248(24):8353–5.
- [Colomb et al., 1969] Colomb, M. G., Chéruey, A., and Vignais, P. V. (1969). Nucleoside diphosphatekinase from beef heart mitochondria: purification and properties. *Biochemistry*, 8(5):1926–39.
- [Crow et al., 1983] Crow, K. E., Braggins, T. J., and Hardman, M. J. (1983). Human liver cytosolic malate dehydrogenase: purification, kinetic properties, and role in ethanol metabolism. *Arch. Biochem. Biophys.*, 225(2):621–9.
- [Davisson and Schulz, 1985] Davisson, V. J. and Schulz, A. R. (1985). The purification and steady-state kinetic behaviour of rabbit heart mitochondrial NAD(P)<sup>+</sup> malic enzyme. *Biochem. J.*, 225(2):335–42.
- [De Rosa et al., 1979] De Rosa, G., Burk, T. L., and Swick, R. W. (1979). Isolation and characterization of mitochondrial alanine aminotransferase from porcine tissue. *Biochim. Biophys. Acta*, 567(1):116–24.
- [Dierks and Krämer, 1988] Dierks, T. and Krämer, R. (1988). Asymmetric orientation of the reconstituted aspartate / glutamate carrier from mitochondria. *Biochim. Biophys. Acta*, 937(1):112–26.
- [Ehrlich et al., 1981] Ehrlich, R. S., Hayman, S., Ramachandran, N., and Colman, R. F. (1981). Re-evaluation of molecular weight of pig heart NAD-specific isocitrate dehydrogenase. *J. Biol. Chem.*, 256(20):10560–4.
- [Fato et al., 1996] Fato, R., Estornell, E., Di Bernardo, S., Pallotti, F., Parenti Castelli, G., and Lenaz, G. (1996). Steady-state kinetics of the reduction of coenzyme Q analogs by complex I (NADH:ubiquinone oxidoreductase) in bovine heart mitochondria and submitochondrial particles. *Biochemistry*, 35(8):2705–16.
- [Fell, 1992] Fell, D. A. (1992). Metabolic control analysis: a survey of its theoretical and experimental development. *Biochem. J.*, 286(Pt 2):313–30.
- [Garces and Cleland, 1969] Garces, E. and Cleland, W. W. (1969). Kinetic studies of yeast nucleoside diphosphate kinase. *Biochemistry*, 8(2):633–40.
- [Grivennikova et al., 1993] Grivennikova, V. G., Gavrikova, E. V., Timoshin, A. A., and Vinogradov, A. D. (1993). Fumarate reductase activity of bovine heart succinate-ubiquinone reductase. new assay system and overall properties of the reaction. *Biochim. Biophys. Acta*, 1140(3):282–92.
- [Guarriero-Bobyleva et al., 1978] Guarriero-Bobyleva, V., Masini, A., Volpi-Becchi, M. A., and Cennamo, C. (1978). Kinetic studies of cytoplasmic and mitochondrial aconitate hydratases from rat liver. *Ital. J. Biochem.*, 27(5):287–99.
- [Hamada et al., 1975] Hamada, M., Koike, K., Nakaula, Y., Hiraoka, T., Koike, M., and Hashimoto, T. (1975). A kinetic study of the  $\alpha$ -keto acid dehydrogenase complexes from pig heart mitochondria. *J. Biochem. (Tokyo)*, 77(5):1047–56.
- [Heckert et al., 1989] Heckert, L. L., Butler, M. H., Reimers, J. M., Albe, K. R., and Wright, B. E. (1989). Purification and characterization of the 2-oxoglutarate dehydrogenase complex from *dictyostelium discoideum*. *J. Gen. Microbiol.*, 135(Pt 1):155–61.

- [Henson and Cleland, 1964] Henson, C. P. and Cleland, W. W. (1964). Kinetic studies of glutamic oxaloacetic transaminase isozymes. *Biochemistry*, 3(3):338–45.
- [Indiveri et al., 1991a] Indiveri, C., Dierks, T., Krämer, R., and Palmieri, F. (1991a). Reaction mechanism of the reconstituted oxoglutarate carrier from bovine heart mitochondria. *Eur. J. Biochem.*, 198(2):339–47.
- [Indiveri et al., 1993] Indiveri, C., Prezioso, G., Dierks, T., Krämer, R., and Palmieri, F. (1993). Kinetic characterization of the reconstituted dicarboxylate carrier from mitochondria: a four-binding-site sequential transport system. *Biochim. Biophys. Acta*, 1143(3):310–8.
- [Indiveri et al., 1994] Indiveri, C., Tonazzi, A., and Palmieri, F. (1994). The reconstituted carnitine carrier from rat liver mitochondria: evidence for a transport mechanism different from that of the other mitochondrial translocators. *Biochim. Biophys. Acta*, 1189(1):65–73.
- [Indiveri et al., 1991b] Indiveri, C., Tonazzi, A., Prezioso, G., and Palmieri, F. (1991b). Kinetic characterization of the reconstituted carnitine carrier from rat liver mitochondria. *Biochim. Biophys. Acta*, 1065(2):231–238.
- [Kholodenko, 1993] Kholodenko, B. N. (1993). Kinetic models of coupling between  $H^+$  and  $Na^+$ -translocation and ATP synthesis/hydrolysis by  $F_oF_1$ -ATPases: Can a cell utilize both  $\Delta\mu_{H^+}$  and  $\Delta\mu_{Na^+}$  for ATP synthesis *in vivo* conditions using the same enzyme? *J. Bioenerg. Biomembr.*, 25(3):285–95.
- [Kiselevsky et al., 1990] Kiselevsky, Y. V., Ostrovtsova, S. A., and Strumilo, S. A. (1990). Kinetic characterization of the pyruvate and oxoglutarate dehydrogenase complexes from human heart. *Acta Biochim. Pol.*, 37(1):135–9.
- [Krämer and Klingenberg, 1982] Krämer, R. and Klingenberg, M. (1982). Electrophoretic control of reconstituted adenine nucleotide translocation. *Biochemistry*, 21(5):1082–9.
- [Kubota et al., 1992] Kubota, T., Yoshikawa, S., and Matsubara, H. (1992). Kinetic mechanism of beef heart ubiquinol:cytochrome *c* oxidoreductase. *J. Biochem. (Tokyo)*, 111(1):91–8.
- [Londesborough and Dalziel, 1970] Londesborough, J. C. and Dalziel, K. (1970). Kinetic studies of NADP-dependent isocitrate dehydrogenase from beef heart mitochondria. In Sund, H., editor, *Pyridine Nucleotide Dependent Dehydrogenases*, pages 315–24. Springer-Verlag, New York.
- [Malmström and Andréasson, 1985] Malmström, B. G. and Andréasson, L. E. (1985). The steady-state rate equation for cytochrome *c* oxidase based on a minimal kinetic scheme. *J. Inorg. Biochem.*, 23(3-4):233–42.
- [Mann et al., 1995] Mann, W. R., Yan, B., Dragland, C. J., and Bell, P. A. (1995). Kinetic, circular dichroism and fluorescence studies on heterologously expressed carnitine palmitoyltransferase II. *J. Enzyme Inhib.*, 9(4):303–8.
- [Matsuno-Yagi and Hatefi, 1985] Matsuno-Yagi, A. and Hatefi, Y. (1985). Studies on the mechanism of oxidative phosphorylation. Catalytic site cooperativity in ATP synthesis. *J. Biol. Chem.*, 260(27):14424–7.
- [Matsuoka and Srere, 1973] Matsuoka, Y. and Srere, P. A. (1973). Kinetic studies of citrate synthase from rat kidney and rat brain. *J. Biol. Chem.*, 248(33):8022–30.
- [McKean et al., 1979] McKean, M. C., Frerman, F. E., and Mielke, D. M. (1979). General acyl-CoA dehydrogenase from pig liver. *J. Biol. Chem.*, 254(8):2730–5.
- [Miyazawa et al., 1981] Miyazawa, S., Furuta, S., Osumi, T., Hashimoto, T., and Ui, N. (1981). Properties of peroxisomal 3-ketoacyl-CoA thiolase from rat liver. *J. Biochem. (Tokyo)*, 90(2):511–9.
- [Mukherjee and Srere, 1976] Mukherjee, A. and Srere, P. A. (1976). Purification of and mechanism studies on citrate synthase. *J. Biol. Chem.*, 251(5):1476–80.
- [Nalecz, 1994] Nalecz, K. A. (1994). The mitochondrial pyruvate carrier: the mechanism of substrate binding. In Forte, M. and Colombini, M., editors, *Molecular Biology of Mitochondrial Transport Systems*, pages 67–79. Springer-Verlag, Berlin, Heidelberg.
- [Plaut et al., 1974] Plaut, G. W. E., Schramm, V. L., and Aogaichi, T. (1974). Action of magnesium ion on diphosphopyridine nucleotide-linked isocitrate dehydrogenase from bovine heart. *J. Biol. Chem.*, 249(6):1848–56.
- [Ramsay et al., 1987] Ramsay, R. R., Derrick, J. P., Friend, A. S., and Tubbs, P. K. (1987). Purification and properties of the soluble carnitine palmitoyltransferase from bovine liver mitochondria. *Biochem. J.*, 244(2):271–8.

- [Shepherd and Garland, 1969] Shepherd, D. and Garland, P. B. (1969). The kinetic properties of citrate synthase from rat liver mitochondria. *Biochem. J.*, 114(3):597–610.
- [Sluse et al., 1991] Sluse, F. E., Evens, A., Dierks, T., Duyckaerts, C., Sluse-Goffart, C. M., and Krämer, R. (1991). Kinetic study of the aspartate / glutamate carrier in intact rat heart mitochondria and comparison with a reconstituted system. *Biochim. Biophys. Acta*, 1058(3):329–38.
- [Stappen and Krämer, 1994] Stappen, R. and Krämer, R. (1994). Kinetic mechanism of phosphate/phosphate and phosphate/ $\text{OH}^-$  antiports catalyzed by reconstituted phosphate carrier from beef heart mitochondria. *J. Biol. Chem.*, 269(15):11240–6.
- [Velick and Vavra, 1962] Velick, S. F. and Vavra, J. (1962). A kinetic and equilibrium analysis of the glutamic oxaloacetate transaminase mechanism. *J. Biol. Chem.*, 237(7):2109–22.
- [Woeltje et al., 1987] Woeltje, K. F., Kuwajima, M., Foster, D. W., and McGarry, J. D. (1987). Characterization of the mitochondrial carnitine palmitoyltransferase enzyme system. *J. Biol. Chem.*, 262(20):9822–7.
- [Yang and Schulz, 1987] Yang, S. Y. and Schulz, H. (1987). Kinetics of coupled enzyme reactions. *Biochemistry*, 26(17):5579–84.